



Prognostic value of multiparametric cardiac magnetic resonance in sickle cell patients

Antonella Meloni^{1,2} · Laura Pistoia¹ · Alessandra Quota³ · Giuseppe Messina⁴ · Paolo Ricchi⁵ · Sergio Bagnato⁶ · Calogera Gerardi⁷ · Roberto Lisi⁸ · Liana Cuccia⁹ · Stefania Renne¹⁰ · Antonino Vallone¹¹ · Riccardo Righi¹² · Vincenzo Positano^{1,2} · Alessia Pepe¹³ · Filippo Cademartiri¹

Received: 20 June 2022 / Accepted: 11 November 2022

© The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2022

Abstract

The aim of this multicenter study was to prospectively assess the predictive value of multiparametric cardiac magnetic resonance (CMR) for cardiovascular complications in sickle cell disease (SCD) patients. Among all patients with hemoglobinopathies consecutively enrolled in the Myocardial Iron Overload in Thalassemia (MIOT) Network, we selected 102 SCD patients (34.38 ± 12.67 years, 49 females). Myocardial iron overload (MIO) was measured by the multislice multiecho T2* technique. Atrial dimensions and biventricular function parameters were quantified by cine images. Late gadolinium enhancement (LGE) images were acquired to detect focal myocardial fibrosis. At baseline CMR, only two patients had significant MIO (global heart T2* < 20 ms). During a mean follow-up of 63.01 ± 24.95 months, 11 cardiovascular events (10.8%) were registered: 3 pulmonary hypertension, 2 supraventricular arrhythmias, 1 heart failure, 1 death for heart failure, 1 pulmonary embolism, 1 peripheral vascular disease, 1 transient ischemic attack, and 1 death after acute chest syndrome. In the multivariate analysis, the independent CMR predictors of cardiovascular events were left ventricular (LV) ejection fraction (hazard ratio-HR = 0.88; $p = 0.025$) and right ventricular (RV) mass index (HR = 1.09; $p = 0.047$). According to the receiver-operating characteristic curve analysis for adverse events, an LV ejection fraction < 58.9% and an RV mass index > 31 g/m² were optimal cut-off values. Reduced left ventricular ejection fraction and increased right ventricular mass index showed a significant prognostic value in patients with SCD. Our data seem to suggest that CMR may be added as a screening tool for identifying SCD patients at high risk for cardiopulmonary and vascular diseases.

Keywords Sickle cell disease · Cardiovascular complications · Magnetic resonance imaging · Prognosis

✉ Filippo Cademartiri
fcademartiri@ftgm.it

¹ Department of Radiology, Fondazione G. Monasterio CNR-Regione Toscana, Via Moruzzi, 1 - 56124 Pisa, Italy

² U.O.C. Bioingegneria, Fondazione G. Monasterio CNR-Regione Toscana, Pisa, Italy

³ Servizio Di Talassemia, Ospedale V. Emanuele III, Gela, CL, Italy

⁴ Centro Microcitemie, Grande Ospedale Metropolitano Bianchi-Melacrino-Morelli, Reggio Calabria, Italy

⁵ U.O.S.D. Malattie Rare del Globulo Rosso, Azienda Ospedaliera Di Rilievo Nazionale "A. Cardarelli", Naples, Italy

⁶ Ematologia Microcitemia, Ospedale San Giovanni di Dio – ASP Crotone, Crotone, Italy

⁷ Unità Operativa Semplice Di Talassemia, Presidio Ospedaliero Giovanni Paolo II - Distretto AG2 Di Sciacca, Sciacca, AG, Italy

⁸ Unità Operativa Dipartimentale Talassemia, Azienda Ospedaliera Garibaldi Presidio Ospedaliero Garibaldi-Centro, Catania, Italy

⁹ Unità Operativa Complessa Ematologia Con Talassemia, ARNAS Civico Benfratelli-Di Cristina, Palermo, Italy

¹⁰ Struttura Complessa Di Cardioradiologia-UTIC, Presidio Ospedaliero "Giovanni Paolo II", Lamezia Terme, Italy

¹¹ Reparto Di Radiologia, Azienda Ospedaliera Garibaldi Presidio Ospedaliero Nesima, Catania, Italy

¹² Diagnostica Per Immagini e Radiologia Interventistica, Ospedale del Delta, Lagosanto, FE, Italy

¹³ Institute of Radiology, Department of Medicine, University of Padua, Padua, Italy

Introduction

Sickle cell disease (SCD) is one of the most common inherited disorders of hemoglobin (Hb) production and has been recognized as a global public health problem. SCD is caused by mutations in the beta-globin gene that lead to the production of an abnormal Hb variant, known as HbS [1]. SCD can occur due to homozygosity for the HbS gene (HbSS), due to compound heterozygosity for HbS and another structural hemoglobin variant, such as hemoglobin C or D, and due to double heterozygosity of HbS and beta-thalassemia (HbS/ β -thalassemia or sickle cell/ β -thalassemia) [2]. HbS/ β -thalassemia represents the most prevalent form of sickling syndromes in Italy due to the high frequency of the β -thalassemia trait.

Thanks to several advances in the diagnosis and treatment, including childhood vaccination, new-borns screening, penicillin prophylaxis for pneumococcal infection in childhood, red blood cell transfusion, hydroxyurea therapy, and comprehensive medical care, early childhood mortality of SCD patients has dramatically decreased in high-income countries. In the aging population, cardiovascular complications are emerging as a major cause of reduced quality of life and early mortality [3, 4].

The two clinical hallmarks of SCD, hemolysis and vaso-occlusive crises with repeated episodes of ischemia and reperfusion, strongly contribute to cardiovascular involvement [5, 6]. The microvascular dysfunction related to repeated vaso-occlusive events and the nitric oxide scavenging resulting from chronic intravascular haemolysis trigger a chronic inflammatory state and widespread vasculopathy, that can damage multiple organs, including the heart and the lungs [7, 8]. The chronic hemolysis-related anemia is associated with a compensatory increase in blood volume [9], which enhances the ventricular pump performance. The anatomical–functional expression of this chronic state is the dilatation of all cardiac chambers [10, 11]. Over time, progressive dilation leads to increased wall stress and eccentric hypertrophy [12]. This volume overload state can lead to increased filling pressures, increased venous return, abnormal pulmonary hemodynamics, arrhythmias, and the syndrome of high-output heart failure [13]. Of note, it has been suggested that the high-output state of SCD, rather than primary abnormalities of the pulmonary microvasculature, represents the major driver of pulmonary hypertension (PH) [14].

Chronic transfusion therapy can reduce both hemolytic anemia and vaso-occlusive sickling episodes [15], but can add another potential factor of stress for the cardiovascular system: a secondary state of iron overload [16]. Cardiac and vascular iron overload may reduce ventricular dimensions initially through vascular and ventricular

stiffening [17, 18] but may increase ventricular dimensions and decrease systolic function in end-stage disease [19, 20]. Myocardial iron overload (MIO) is relatively rare in patients with SCD [21–23], but the increasing life expectancy and duration of chronic transfusion will make MIO a more significant clinical problem.

Due to its multiparametric nature, cardiac magnetic resonance (CMR) represents a powerful tool to evaluate structural and functional impairments in the myocardium of SCD patients. T2* CMR is the method of choice for the non-invasive, fast, and reproducible quantification of MIO [24] and has been validated against histological findings [25, 26]. CMR is the gold standard for the non-invasive assessment of biventricular size and function with excellent accuracy and reproducibility [27]. In particular, CMR provides the most comprehensive information on the right ventricle, by virtue of its high spatial and temporal resolution, its excellent signal-to-noise ratio between the myocardium and the blood pool, and the fact that, conversely to echocardiography, it is free from acoustic window limitations and independent of geometric assumptions. Finally, following the injection of a contrast agent, CMR represents a valuable tool for the detection of myocardial fibrosis [28]. In SCD, many processes including anemia, ischemia, inflammation, and microvascular disease may predispose to myocardial fibrosis [29].

There are no prospective cohort studies evaluating the association between multiparametric CMR findings (heart iron, function, and fibrosis) and cardiovascular outcomes in SCD patients. Therefore, the aim of this multicenter study was to prospectively assess the predictive value of CMR parameters for cardiovascular complications in SCD patients.

Methods

Patients

Among all patients with hemoglobinopathies consecutively enrolled in the Myocardial Iron Overload in Thalassemia (MIOT) Network, we selected only those with SCD ($N=109$: 48% females, mean age 35.08 ± 12.87 years). Globally, the following inclusion criteria were adopted in the MIOT Network: (1) male and female patients, of all ages, with thalassemia syndromes or structural hemoglobin variants, requiring magnetic resonance imaging (MRI) to quantify cardiac and liver iron burden; (2) written informed consent; (3) written authorization for use and disclosure of protected health information; (4) no absolute contraindications to MRI.

The MIOT Network was a collaborative project among more than 60 hematological centers and 10 validated MRI sites, where MRI exams were performed using

homogeneous, standardized, and validated procedures [30, 31]. All centers were linked by a shared database [32], where the clinical-anamnestic history of the patients, from birth to the date of the first MRI scan, was recorded. All patients performed a routine screening and at every MRI follow-up, performed by protocol every 18 ± 3 months, the clinical, instrumental, and laboratory data were updated. Clinical follow-up continued until September 2016. Each hematologist completed a case report form detailing patient outcomes between the last MRI and September 2016.

The study complied with the Declaration of Helsinki and was approved by the institutional ethics committee. All patients provided informed consent.

Magnetic resonance

All MRI examinations were performed with a clinical 1.5 T scanner (GE Healthcare, Milwaukee, WI, USA). An eight-element cardiac phased-array receiver surface coil with breath-holding in end-expiration and ECG-gating was used.

The T2* technique was used for iron overload assessment. A single mid-hepatic slice [33] and three parallel short-axis views (basal, medium, and apical) of the left ventricle (LV) were acquired in the same imaging session [30, 34]. T2* images analysis was performed using a custom-written, previously validated software (HIPPIOT®) [35]. Hepatic T2* values were calculated in a circular region of interest [36] and were converted into liver iron concentration (LIC) using Wood's calibration curve [37, 38]. The software provided the T2* value for all 16 segments of the LV, according to the standard American Heart Association (AHA)/American College of Cardiology (ACC) model [39], and the global heart T2* value was obtained by averaging all segmental values.

For the quantification of biventricular function parameters, cine images were acquired in sequential 8 mm short-axis slices (gap 0 mm) from the atrioventricular ring to the apex. Images were analyzed in a standard way using MASS® software (Medis, Leiden, The Netherlands) [40]. Atrial areas were measured from the 4-chamber view projection in the ventricular end-systolic phase. Biventricular volumes and masses and bi-atrial areas were normalized for the body surface area.

To detect the presence of focal/macrosopic myocardial fibrosis, late gadolinium enhancement (LGE) short-axis and vertical, horizontal, and oblique long-axis images were acquired 10–18 min after Gadobutrol (Gadovist®; Bayer Schering Pharma; Berlin, Germany) intravenous administration at the standard dose of 0.2 mmol/kg. LGE images were not acquired in patients with a glomerular filtration rate < 30 mL/min/1.73m² and in patients who refused. LGE was considered present when visualized in two different views [41, 42].

Diagnostic criteria

An MR LIC ≥ 3 mg/g/dw was considered indicative of significant iron load [43]. A T2* measurement of 20 ms was taken as a “conservative” normal value for segmental and global values [20].

The outcome of this study was the incidence of cardiovascular complications, defined as a composite of cardiac complications and pulmonary, cerebral, and peripheral vascular diseases. Heart failure (HF) was identified based on symptoms, signs, biomarkers, and instrumental parameters, according to the current guidelines [44]. Arrhythmias were diagnosed and classified according to the AHA/ACC Guidelines [45]. PH was diagnosed if the trans-tricuspidal velocity jet on trans-thoracic echocardiogram was > 3.2 m/s [46] in presence of signs and symptoms. In case of suspicion, the diagnosis of pulmonary embolism (PE) was accurately confirmed or ruled out by non-invasive imaging tests [47]. The diagnosis of a transient ischemic attack (TIA) was made based on symptoms, objective findings on neurologic examination, and imaging of the brain [48]. The clinical diagnosis of deep vein thrombosis was confirmed by objective testing using ultrasound or venography. If a patient developed more than one complication, only the first one was considered.

Statistical analysis

All data were analyzed using SPSS version 27.0 (IBM Corp, Armonk, NY) and MedCalc version 19.8 (MedCalc Software Ltd, Ostend, Belgium) statistical packages.

Continuous variables were described as mean \pm standard deviation (SD). Categorical variables were expressed as frequencies and percentages.

The normality of the distribution of the continuous variables was assessed by using the Kolmogorov–Smirnov test.

Comparisons between two groups were made by independent-samples *t*-test for continuous variables with normal distribution and Mann–Whitney *U* test for continuous variables with non-normal distribution. χ^2 testing was performed for categorical variables.

Correlation analysis was performed using Pearson's test or Spearman's test where appropriate.

The Cox proportional-hazard model was used to test the association between the considered prognostic variables and the outcome. The variables with a statistical significance in the univariable analysis were placed in the multivariate model. They were ruled out if they did not significantly improve the adjustment of the model. The results were presented as hazard ratio (HR) with 95% confidence intervals (CI).

The optimal cut-off value of clinical variables with statistical significance in the multivariable analysis was assessed

using a receiver-operating characteristic (ROC) curve analysis for the endpoint of this study.

A $P < 0.05$ was considered statistically significant.

Results

Baseline data

Seven patients were excluded from this study because a cardiac complication (4 arrhythmias, 2 HF, and 1 PH) was present at the baseline MRI.

The baseline demographic, clinical, and MRI features of the 102 considered SCD patients are described in Table 1. Patients were homogeneously distributed in terms of gender and the mean age was 34.38 ± 12.67 years. Forty-nine (48.0%) patients were under regular transfusion regimen (≥ 4 transfusions/year): 39 received simple transfusions (mean number of transfusions/year 11.78 ± 6.62) and 10 exchange transfusions (mean number of transfusions/year 10.50 ± 3.93). No significant difference in serum hemoglobin levels was detected between never/sporadically transfused patients and regularly transfused patients (9.44 ± 1.25 g/dl vs 9.42 ± 1.24 g/dl; $P = 0.881$). Hemoglobin levels were not

associated with biventricular volume or mass indexes but showed a weak positive correlation with LV ejection fraction (EF) ($R = 0.232$; $P = 0.029$). Half of SCD patients were on no iron chelation therapy. Among the 51 chelated patients, 27 were receiving deferoxamine, 15 deferasirox, 7 deferriprone, and 2 deferoxamine in combination with deferriprone. Hepatic and myocardial iron overload were detected, respectively, in the 54.9% (56/102) and 2.0% (2/102) of patients. The contrast medium was administered only in 66 patients, of whom 10 (15.2%) had nonischemic focal myocardial fibrosis. A mesocardial LGE at the insertion points of the free RV wall in the interventricular septum was found in 5 patients.

Twenty-four patients had homozygous HbSS and 78 patients had HbS/ β -thalassemia. The latter group included both HbS/ $\beta 0$ thalassemia and HbS/ $\beta +$ thalassemia patients, not further differentiated according to the beta-globin mutation. The comparison between HbSS and HbS/ β -thalassemia groups is shown in Table 1. Surgical splenectomy and treatment with hydroxyurea were significantly less common in HBSS than in HbS/ β -thalassemia patients, but HBSS patients were more frequently regularly transfused. Hepatic and cardiac iron overload, biventricular function parameters, and bi-atrial areas were comparable between the two groups.

Table 1 Baseline demographic, clinical and MRI findings in SCD patients divided into two groups based on the SCD genotype

Variable	All patients (N=102)	Homozygous HbS (N=24)	Hbs/ β -thalassemia (N=78)	P
Females, N (%)	49 (48.0)	10 (41.7)	39 (50.0)	0.475
Age (years)	34.38 ± 12.67	30.84 ± 12.49	35.47 ± 12.61	0.117
Splenectomy, N (%)	58 (56.9)	7 (29.2)	51 (65.4)	0.002
Regularly transfusions, N (%)	49 (48.0)	17 (70.8)	32 (41.0)	0.011
Chelation therapy, N (%)	51 (50.0)	12 (50.0)	39 (50.0)	1.000
Hydroxyurea therapy, N (%)	48/85 (56.5)	8/21 (38.1)	40/64 (62.5)	0.050
Serum hemoglobin (g/dl)	9.43 ± 1.24	9.72 ± 1.40	9.35 ± 1.19	0.267
Serum ferritin (ng/l)	1308.99 ± 1528.83	1714.42 ± 1824.36	1200.49 ± 1435.04	0.308
MRI LIC (mg/g/dw)	6.82 ± 9.89	9.22 ± 16.79	6.08 ± 6.45	0.862
MRI LIC ≥ 3 mg/g/dw, N (%)	56 (54.9)	13 (54.2)	43 (55.1)	0.934
Global heart T2*(ms)	35.99 ± 6.42	37.99 ± 5.12	35.37 ± 6.68	0.080
Global heart T2* < 20 ms, N (%)	2 (2.0)	0 (0.0)	2 (2.6)	1.000
N. of segments with T2* < 20 ms	1.06 ± 2.37	0.46 ± 1.14	1.24 ± 2.61	0.069
LV EDVI (ml/m ²)	93.71 ± 20.73	94.35 ± 22.39	93.51 ± 20.33	0.806
LV mass index (g/m ²)	62.17 ± 16.95	63.24 ± 21.89	61.83 ± 15.23	0.859
LV EF (%)	61.39 ± 6.86	63.35 ± 5.82	60.78 ± 7.07	0.109
RV EDVI (ml/m ²)	84.17 ± 20.51	83.92 ± 20.92	84.25 ± 20.52	0.732
RV mass index (g/m ²)	30.31 ± 8.98	34.58 ± 11.45	29.91 ± 7.63	0.070
RV EF (%)	63.13 ± 7.82	63.42 ± 8.18	63.04 ± 7.76	0.839
Focal myocardial fibrosis, N(%)	10/66 (15.2)	3/15 (20.0)	7/51 (13.7)	0.683
Left atrial area (cm ² /m ²)	12.77 ± 2.61	13.62 ± 3.16	12.41 ± 2.28	0.089
Right atrial area (cm ² /m ²)	11.89 ± 2.26	11.69 ± 2.38	11.99 ± 2.22	0.640

N, number; MRI, magnetic resonance imaging; LIC, liver iron concentration; LV, left ventricular; EDVI, end-diastolic volume index; EF, ejection fraction; RV, right ventricular

Prediction of cardiovascular complications

The mean follow-up time was 63.01 ± 24.95 months (median = 64.71 months). Cardiovascular events were recorded in 11 (10.8%) patients: 3 pulmonary hypertension, 2 supraventricular arrhythmias, 1 heart failure, 1 death for heart failure, 1 pulmonary embolism, 1 peripheral vascular disease, 1 transient ischemic attack, and 1 death after acute chest syndrome. The mean age at the first complication was 45.86 ± 10.21 years (range: 29–57 years). The mean time from the first MRI to the development of a cardiac complication was 50.77 ± 26.72 months.

Table 2 shows the comparison of baseline characteristics as well as MRI parameters between patients free of events and patients who developed a cardiovascular event. No significant difference was detected for gender, type of SCD (HbS homozygosity vs compound heterozygosity for HbS and either $\beta 0$ or $\beta +$ thalassemia), presence of regular transfusions or chelation therapy, and indices of iron overload. Cardiovascular events were associated with aging and with lower baseline serum hemoglobin levels. Patients suffering a cardiovascular event had significantly lower LV EF and

significantly higher right ventricular (RV) mass index at the baseline MRI while no difference was found in all other biventricular function parameters or atrial areas.

Table 3 shows the results of the univariate Cox regression analysis. Among the non-MRI parameters, aging and lower serum hemoglobin levels emerged as the significant univariate prognosticators of cardiovascular complications. Multivariate analysis revealed that both variables remained prognostic indicators (age: HR = 1.08, 95%CI = 1.01–1.15, $P = 0.025$ and serum hemoglobin: HR = 0.33, 95%CI = 0.14–0.76, $P = 0.010$). No significant correlation was detected between age and serum hemoglobin levels ($R = -0.064$, $P = 0.551$). Among the MRI parameters, LV EF and RV mass index were significant univariate prognosticators of cardiovascular complications. Both variables remained significant prognosticators at the multivariate analysis (LV EF: HR = 0.88, 95%CI = 0.79–0.98, $P = 0.025$ and RV mass index: HR = 1.09, 95%CI = 1.01–1.18, $P = 0.047$). Due to the low number of events, it was not possible to perform a multivariate model including all four univariate prognosticators.

The patient who died of HF showed a baseline global heart $T2^* = 9.94$ ms and all segments with $T2^* < 20$ ms.

Table 2 Comparison of baseline characteristics in SCD patients free of events versus those who developed a cardiovascular event during the follow-up

Variable	No cardiovascular events ($N = 91$)	Cardiovascular events ($N = 11$)	P
Females, N (%)	47 (51.6)	6 (54.5)	0.856
Age (yrs)	33.51 ± 12.69	41.63 ± 10.37	0.044
Type of SCD, N (%)			
Homozygous HbS	23 (25.3)	1 (9.1)	0.451
HbS/ β -thalassemia	68 (74.7)	10 (90.9)	
Splenectomy, N (%)	51 (56.0)	7 (63.6)	0.753
Regular transfusions, N (%)	44 (48.4)	5 (45.5)	0.856
Chelation therapy, N (%)	44 (48.4)	7 (63.9)	0.525
Hydroxyurea therapy, N (%)	41/75 (54.7)	7/10 (70.0)	0.502
Serum hemoglobin (g/dl)	9.56 ± 1.20	8.41 ± 1.07	0.001
Serum ferritin (ng/l)	1323.62 ± 1564.33	1191.90 ± 1271.25	0.653
MRI LIC (mg/g/dw)	6.83 ± 10.19	6.72 ± 7.28	0.750
MRI LIC ≥ 3 mg/g/dw, N (%)	49 (53.8)	7 (63.6)	0.750
Global heart $T2^*$ (ms)	36.11 ± 5.90	35.00 ± 10.11	0.817
Global heart $T2^* < 20$ ms, N (%)	1 (1.1)	9 (9.1)	0.205
N . of segments with $T2^* < 20$ ms	0.95 ± 1.91	2.00 ± 4.79	0.955
LV EDVI (ml/m^2)	93.36 ± 20.76	96.53 ± 21.28	0.559
LV mass index (g/m^2)	61.54 ± 16.83	67.25 ± 17.89	0.268
LV EF (%)	62.01 ± 6.72	56.47 ± 6.13	0.011
RV EDVI (ml/m^2)	83.99 ± 20.58	85.65 ± 20.85	0.804
RV mass index (g/m^2)	29.71 ± 9.22	34.87 ± 5.23	0.031
RV EF (%)	63.50 ± 7.55	60.13 ± 9.67	0.179
Focal myocardial fibrosis, N (%)	10/59 (16.9)	0/7 (0.0)	0.583
Left atrial area (cm^2/m^2)	12.84 ± 2.65	12.13 ± 2.27	0.500
Right atrial area (cm^2/m^2)	11.85 ± 2.31	12.29 ± 1.89	0.627

N , number; SCD, sickle cell disease; MRI, magnetic resonance imaging; LIC, liver iron concentration; LV, left ventricular; EDVI, end-diastolic volume index; EF, ejection fraction; RV, right ventricular

Table 3 Results of univariate and multivariate Cox regression analysis

	Univariate analysis	
	HR (95%CI)	P
Male gender	0.95 (0.29–3.15)	0.933
Age	1.07 (1.01–1.13)	0.034
Homozygous HBS mutation	0.29 (0.04–2.27)	0.237
Splenectomy	1.20 (0.35–4.13)	0.768
Regularly transfusions	0.78 (0.23–2.62)	0.686
Chelation therapy	1.66 (0.49–5.71)	0.418
Hydroxyurea therapy	1.42 (0.36–5.62)	0.615
Serum hemoglobin	0.034 (0.16–0.74)	0.006
Serum ferritin	1.00 (1.00–1.00)	0.780
MRI LIC	1.00 (0.95–1.06)	0.941
Global heart T2*	0.98 (0.89–1.07)	0.631
N. of segments with T2* < 20 ms	1.12 (0.94–1.34)	0.188
LV EDVI	1.01 (0.98–1.03)	0.827
LV mass index	1.02 (0.98–1.05)	0.402
LV EF	0.88 (0.80–0.97)	0.007
RV EDVI	1.00 (0.98–1.03)	0.876
RV mass index	1.07 (1.01–1.14)	0.046
RV EF	0.94 (0.87–1.03)	0.175
Focal myocardial fibrosis	0.04 (0.00–48.10)	0.501
Left atrial area index	0.88 (0.66–1.18)	0.384
Right atrial area index	1.01 (0.74–1.39)	0.936

N, number; SCD, sickle cell disease; MRI, magnetic resonance imaging; LIC, liver iron concentration; LV, left ventricular; EDVI, end-diastolic volume index; EF, ejection fraction; RV, right ventricular

The patient who developed HF had a baseline global heart T2* = 24.94 ms but 4 segments with T2* < 20 ms. The other patient with a baseline global heart T2* < 20 ms did not develop a cardiovascular complication during the follow-up but after the MRI she changed the chelation regimen, switching from deferoxamine in monotherapy to sequential deferoxamine/deferiprone.

Optimal cut-off values of CMR predictors for cardiovascular complications

At ROC curve analysis, an LV EF < 58.9% predicted the presence of future cardiovascular events with a sensitivity of 72.7 and a specificity of 71.9 ($P = 0.002$). The area under the curve was 0.71 (95%CI: 0.63–0.81) (Fig. 1A).

A RV mass index > 31 g/m² predicted the presence of future cardiovascular events with a sensitivity of 87.5%

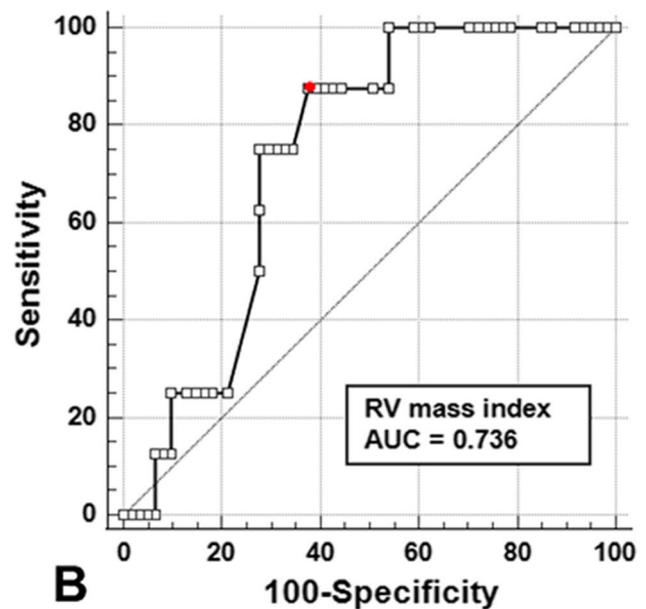
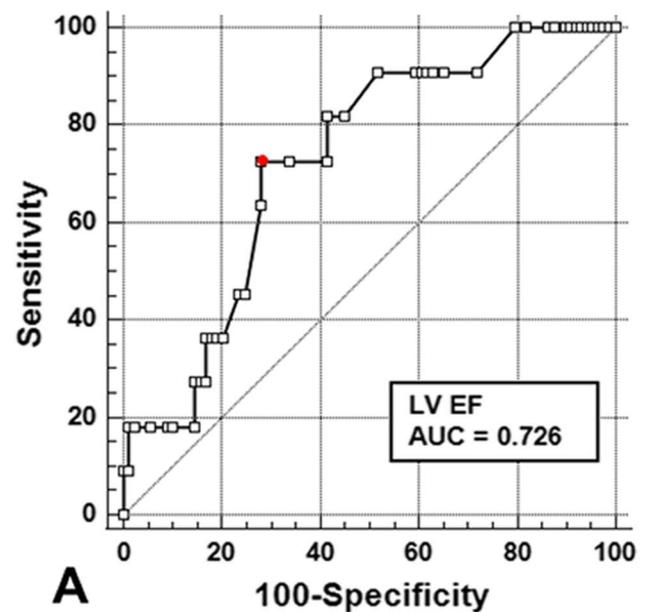


Fig. 1 ROC curve analysis of left ventricular ejection fraction (A) and RV mass index (B) to predict cardiovascular events

and a specificity of 62.3% ($P = 0.001$). The area under the curve was 0.74 (95%CI: 0.62–0.84) (Fig. 1B).

Discussion

To the best of our knowledge, this is the first study exploring the value of multiparametric CMR, including RV mass assessment, in the prognostic evaluation of SCD patients.

We included patients with homozygous HbS and HbS/ β -thalassemia. HbS/ β -thalassemias are classified as HbS/ β 0 thalassemia and HbS/ β + thalassemia, with the former being characterized by the absence of adult hemoglobin and a severe clinical course similar to homozygous SCD [49]. HbS/ β + thalassemia is clinically heterogeneous, with variable residual amounts of adult hemoglobin that determine the clinical course [50]. A recent echocardiographic study showed that diameters, thicknesses, masses, and volumes of cardiac chambers were comparable between HbS/ β 0 thalassemia and HbS/ β + thalassemia patients, suggesting that, unlike other clinical parameters, the cardiac involvement in this disease does not depend so much on the thalassemia genotype [51]. In the present study, we detected comparable cardiac T2* values and biventricular function parameters by CMR between homozygous HbS and HbS/ β -thalassemia patients and we considered all patients as a unique group, irrespective of the genotype.

In our cohort of SCD patients, reduced LV EF and increased RV mass index emerged as CMR-independent predictors of cardiovascular complications. LV EF is the most commonly used surrogate marker of LV systolic function. A meta-analysis including 19 studies reported no significant differences between SCD patients and controls in LV EF assessed by echocardiography [52]. Unlike diastolic dysfunction, systolic dysfunction is considered rare among SCD patients [53]. Indeed, according to our ROC analysis, a relatively high threshold for LV EF (<58.9%) predicted the presence of future cardiovascular events. In our cohort, LV EF was significantly correlated with hemoglobin, suggesting that chronic anemia plays a key role in depressing LV systolic function and, consequently, in the development of cardiovascular complications. In support of this thesis, serum hemoglobin emerged as a clinical univariate prognosticator of cardiovascular complications. However, the microcirculation damage due to vaso-occlusive crisis has been demonstrated to be another important contributor to the deterioration of LV systolic function [54]. In SCD, abnormal myocardial perfusion and flow reserve, related to erythrocyte sickling that occludes the small arteries, capillaries, and venules and endothelial proliferation, have been demonstrated by echocardiography [54] and myocardial scintigraphy [55, 56]. In a study involving 22 children with SCD, myocardial perfusion defects were found in 8 patients, of whom 5 had cardiac symptoms (three episodes of cardiac failure, one of ventricular fibrillation, and one angina) [56].

Traditionally, the importance of the RV has been underestimated and overlooked in clinical practice and literature. However, in the last decades, the central role of the RV in the management and prognosis of many cardiac diseases has been recognized, changing our perspective towards the right side of the heart [57]. Junqueira et al. showed that SCD patients had significantly higher RV

mass index than healthy subjects [58]. Pulmonary vascular endothelial damage/dysfunction can result in the loss of vascular reactivity, and activation of proliferative and antiapoptotic pathways, leading to vascular remodeling, and elevated pulmonary artery pressure [59]. Moreover, the high cardiac output causes increased pulmonary pressure regardless of whether pulmonary vascular resistance is high or not [60]. As pulmonary pressures increase, the thin-walled RV begins to hypertrophy and based on our results, an RV mass index of $31 > \text{g/m}^2$ can predict the development of cardiovascular events. So, although larger studies are needed to confirm the prognostic impact of an increased RV mass index, the RV mass assessment should be included in the routine MRI of SCD patients. Since all three patients who developed PH had an RV mass index $> 31 \text{ g/m}^2$, the combination of both Doppler echocardiography, which represents the non-invasive screening test for PH [61], and MRI may increase the positive predictive value [62] for the detection of right heart catheterization (RHC)-confirmed PH. Although trans-thoracic echocardiography is largely used as an initial imaging modality, it has limited diagnostic capabilities for the evaluation of RV due to its thin wall, peculiar morphology, and the anterior position in the chest.

In contrast to thalassemia major patients [63, 64], we did not detect an association between MIO and cardiac complications, most likely because significant MIO was detected only in two patients, and there were only two cases of heart failure. It has been demonstrated that MIO contributes less to the development of arrhythmias and pulmonary hypertension than cardiac failure [63–66]. However, it should be pointed out that one of the two patients with significant MIO died of HF and in the other patient, the abnormal T2* prompted changes in clinical management. Indeed, the early start of aggressive chelation therapy can prevent, delay, or even reverse iron cardiomyopathy [67]. Although SCD patients have a lower risk for developing myocardial siderosis as compared to other hemoglobinopathies [68], our findings suggest once cardiac iron is present, it is associated with its own toxicity.

The contrast medium was administered only in 66% of patients, partially explaining the absence of a correlation between focal fibrosis and cardiovascular complications. Most importantly, the LGE technique relies on the contrast between normal and abnormal myocardium areas and cannot accurately detect a more diffuse fibrotic process affecting the whole myocardium [69]. Conversely to focal fibrosis [58], diffuse myocardial fibrosis was found to be a common process in both mice models [70] and patients with SCD [71], associated with diastolic dysfunction and the restrictive physiology features of SCD-related cardiomyopathy.

Beyond CMR, aging emerged as a significant predictor of cardiovascular complications. Aging itself results in

well-defined phenotypic changes, which render the cardiovascular system prone to disease, even in the absence of traditional and non-traditional risk factors [72]. On the other hand, as patients live longer, the chronic effects of sustained hemolytic anemia and vaso-occlusive events accumulate [53].

This study is limited by the low number of patients and of cardiovascular events, which prevented us to evaluate the prognostic association between the MRI parameters and the different types of cardiovascular events, considered separately. Larger studies are needed to explore this issue. Another limitation is that the diagnosis of PH was not confirmed by RHC. Although RHC is the diagnostic gold standard for PH [73], it is an invasive and expensive procedure, unsuitable as a screening tool. Moreover, we did not have sufficient information on the quantitative β -globin defect of HbS/ β -thalassemia patients ($\beta 0$ or $\beta +$ mutation) and we did not assess at the baseline MRI the hemoglobin A percentage, which could have been useful to better characterize or stratify our population.

Conclusion

Reduced left ventricular ejection fraction and increased right ventricular mass index showed a significant prognostic value in patients with SCD. This finding suggests that multiparametric CMR may be added as a screening tool for identifying SCD patients at high risk for cardiopulmonary and vascular diseases.

Acknowledgements We would like to thank all the colleagues involved in the MIOT project and all patients for their cooperation.

Funding The MIOT project received “no-profit support” from Chiesi Farmaceutici S.p.A.

Data availability The datasets generated during and/or analysed during the current study are available from the corresponding author on reasonable request.

Declarations

Conflict of interest The authors declare no competing interests.

References

- Ashley-Koch A, Yang Q, Olney RS (2000) Sick cell hemoglobin (HbS) allele and sickle cell disease: a HuGE review. *Am J Epidemiol* 151(9):839–845. <https://doi.org/10.1093/oxfordjournals.aje.a010288>
- Steinberg MH (2008) Sick cell anemia, the first molecular disease: overview of molecular etiology, pathophysiology, and therapeutic approaches. *ScientificWorldJournal* 8:1295–1324. <https://doi.org/10.1100/tsw.2008.157>
- Lanzkron S, Carroll CP, Haywood C Jr (2013) Mortality rates and age at death from sickle cell disease: U.S., 1979–2005. *Public Health Rep* 128(2):110–116. <https://doi.org/10.1177/003335491312800206>
- Voskaridou E, Christoulas D, Terpos E (2012) Sick cell disease and the heart: review of the current literature. *Br J Haematol* 157(6):664–673. <https://doi.org/10.1111/j.1365-2141.2012.09143.x>
- Sachdev V, Rosing DR, Thein SL (2021) Cardiovascular complications of sickle cell disease. *Trends Cardiovasc Med* 31(3):187–193. <https://doi.org/10.1016/j.tcm.2020.02.002>
- Gladwin MT, Sachdev V (2012) Cardiovascular abnormalities in sickle cell disease. *J Am Coll Cardiol* 59(13):1123–1133. <https://doi.org/10.1016/j.jacc.2011.10.900>
- Kato GJ, Hebbel RP, Steinberg MH, Gladwin MT (2009) Vasculopathy in sickle cell disease: biology, pathophysiology, genetics, translational medicine, and new research directions. *Am J Hematol* 84(9):618–625. <https://doi.org/10.1002/ajh.21475>
- Hebbel RP, Belcher JD, Vercellotti GM (2020) The multifaceted role of ischemia/reperfusion in sickle cell anemia. *J Clin Invest* 130(3):1062–1072. <https://doi.org/10.1172/JCI133639>
- Varat MA, Adolph RJ, Fowler NO (1972) Cardiovascular effects of anemia. *Am Heart J* 83(3):415–426
- Lindsay J Jr, Meshel JC, Patterson RH (1974) The cardiovascular manifestations of sickle cell disease. *Arch Intern Med* 133(4):643–651
- Kremastinos DT, Tsiapras DP, Tsetsos GA, Rentoukas EI, Vrettou HP, Toutouzas PK (1993) Left ventricular diastolic Doppler characteristics in beta-thalassemia major. *Circulation* 88(3):1127–1135
- Grossman W, Jones D, McLaurin LP (1975) Wall stress and patterns of hypertrophy in the human left ventricle. *J Clin Invest* 56(1):56–64. <https://doi.org/10.1172/JCI108079>
- Reddy YNV, Borlaug BA (2016) High-output heart failure in sickle cell anemia. *JACC Cardiovasc Imaging* 9(9):1122–1123. <https://doi.org/10.1016/j.jcmg.2016.04.004>
- Mushemi-Blake S, Melikian N, Drasar E, Bhan A, Lunt A, Desai SR, Greenough A, Monaghan MJ, Thein SL, Shah AM (2015) Pulmonary haemodynamics in sickle cell disease are driven predominantly by a high-output state rather than elevated pulmonary vascular resistance: a prospective 3-dimensional echocardiography/Doppler study. *PLoS ONE* 10(8):e0135472. <https://doi.org/10.1371/journal.pone.0135472>
- Ware R (2007) Principles and indications of chronic transfusion therapy for children with sickle cell disease. *Clin Adv Hematol Oncol* 5(9):686–688
- Gordeuk VR, Bacon BR, Brittenham GM (1987) Iron overload: causes and consequences. *Annu Rev Nutr* 7:485–508
- Li W, Coates T, Wood JC (2008) Atrial dysfunction as a marker of iron cardiotoxicity in thalassemia major. *Haematologica* 93(2):311–312
- Cheung YF, Chan GC, Ha SY (2008) Effect of deferasirox (ICL670) on arterial function in patients with beta-thalassaemia major. *Br J Haematol* 141(5):728–733
- Wood JC, Enriquez C, Ghugre N, Otto-Duessel M, Aguilar M, Nelson MD, Moats R, Coates TD (2005) Physiology and pathophysiology of iron cardiomyopathy in thalassemia. *Ann N Y Acad Sci* 1054:386–395
- Anderson LJ, Holden S, Davis B, Prescott E, Charrier CC, Bunce NH, Firmin DN, Wonke B, Porter J, Walker JM, Pennell DJ (2001) Cardiovascular T2-star (T2*) magnetic resonance for the early diagnosis of myocardial iron overload. *Eur Heart J* 22(23):2171–2179
- Wood JC, Tyszka JM, Carson S, Nelson MD, Coates TD (2004) Myocardial iron loading in transfusion-dependent thalassemia and sickle cell disease. *Blood* 103(5):1934–1936
- Meloni A, Puliyl M, Pepe A, Berdoukas V, Coates TD, Wood JC (2014) Cardiac iron overload in sickle-cell disease. *Am J Hematol* 89(7):678–683. <https://doi.org/10.1002/ajh.23721>

23. Tavares AHJ, Benites BD, Fertrin KY (2019) Myocardial iron overload in sickle cell disease: a rare but potentially fatal complication of transfusion. *Transfus Med Rev* 33(3):170–175. <https://doi.org/10.1016/j.tmr.2019.04.001>
24. Wood JC (2011) Impact of iron assessment by MRI. *Hematology Am Soc Hematol Educ Program* 2011:443–450
25. Carpenter JP, He T, Kirk P, Roughton M, Anderson LJ, de Noronha SV, Sheppard MN, Porter JB, Walker JM, Wood JC, Galanello R, Forni G, Catani G, Matta G, Fucharoen S, Fleming A, House MJ, Black G, Firmin DN, St Pierre TG, Pennell DJ (2011) On T2* magnetic resonance and cardiac iron. *Circulation* 123(14):1519–1528
26. Meloni A, Maggio A, Positano V, Leto F, Angelini A, Putti MC, Maresi E, Pucci A, Basso C, Perazzolo Marra M, Pistoia L, De Marchi D, Pepe A (2020) CMR for myocardial iron overload quantification: calibration curve from the MIOT network. *Eur Radiol* 29(5):2246–2252
27. Grothues F, Moon JC, Bellenger NG, Smith GS, Klein HU, Pennell DJ (2004) Interstudy reproducibility of right ventricular volumes, function, and mass with cardiovascular magnetic resonance. *Am Heart J* 147(2):218–223. <https://doi.org/10.1016/j.ahj.2003.10.005>
28. Wu E, Judd RM, Vargas JD, Klocke FJ, Bonow RO, Kim RJ (2001) Visualisation of presence, location, and transmural extent of healed Q-wave and non-Q-wave myocardial infarction. *Lancet* 357(9249):21–28
29. Wood JC (2016) The heart in sickle cell disease, a model for heart failure with preserved ejection fraction. *Proc Natl Acad Sci U S A* 113(35):9670–9672. <https://doi.org/10.1073/pnas.1611899113>
30. Pepe A, Positano V, Santarelli F, Sorrentino F, Cracolici E, De Marchi D, Maggio A, Midiri M, Landini L, Lombardi M (2006) Multislice multiecho T2* cardiovascular magnetic resonance for detection of the heterogeneous distribution of myocardial iron overload. *J Magn Reson Imaging* 23(5):662–668
31. Ramazzotti A, Pepe A, Positano V, Rossi G, De Marchi D, Brizi MG, Luciani A, Midiri M, Sallustio G, Valeri G, Caruso V, Centra M, Cianciulli P, De Sanctis V, Maggio A, Lombardi M (2009) Multicenter validation of the magnetic resonance T2* technique for segmental and global quantification of myocardial iron. *J Magn Reson Imaging* 30(1):62–68
32. Meloni A, Ramazzotti A, Positano V, Salvatori C, Mangione M, Marcheschi P, Favilli B, De Marchi D, Prato S, Pepe A, Sallustio G, Centra M, Santarelli MF, Lombardi M, Landini L (2009) Evaluation of a web-based network for reproducible T2* MRI assessment of iron overload in thalassemia. *Int J Med Inform* 78(8):503–512
33. Positano V, Salani B, Pepe A, Santarelli MF, De Marchi D, Ramazzotti A, Favilli B, Cracolici E, Midiri M, Cianciulli P, Lombardi M, Landini L (2009) Improved T2* assessment in liver iron overload by magnetic resonance imaging. *Magn Reson Imaging* 27(2):188–197
34. Meloni A, Positano V, Pepe A, Rossi G, Dell’Amico M, Salvatori C, Keilberg P, Filosa A, Sallustio G, Midiri M, D’Ascola D, Santarelli MF, Lombardi M (2010) Preferential patterns of myocardial iron overload by multislice multiecho T2* CMR in thalassemia major patients. *Magn Reson Med* 64(1):211–219
35. Positano V, Pepe A, Santarelli MF, Scattini B, De Marchi D, Ramazzotti A, Forni G, Borgna-Pignatti C, Lai ME, Midiri M, Maggio A, Lombardi M, Landini L (2007) Standardized T2* map of normal human heart in vivo to correct T2* segmental artefacts. *NMR Biomed* 20(6):578–590
36. Meloni A, Luciani A, Positano V, De Marchi D, Valeri G, Restaino G, Cracolici E, Caruso V, Dell’Amico MC, Favilli B, Lombardi M, Pepe A (2011) Single region of interest versus multislice T2* MRI approach for the quantification of hepatic iron overload. *J Magn Reson Imaging* 33(2):348–355
37. Wood JC, Enriquez C, Ghugre N, Tyzka JM, Carson S, Nelson MD, Coates TD (2005) MRI R2 and R2* mapping accurately estimates hepatic iron concentration in transfusion-dependent thalassemia and sickle cell disease patients. *Blood* 106(4):1460–1465
38. Meloni A, Rienhoff HY Jr, Jones A, Pepe A, Lombardi M, Wood JC (2013) The use of appropriate calibration curves corrects for systematic differences in liver R2* values measured using different software packages. *Br J Haematol* 161(6):888–891
39. Cerqueira MD, Weissman NJ, Dilsizian V, Jacobs AK, Kaul S, Laskey WK, Pennell DJ, Rumberger JA, Ryan T, Verani MS (2002) Standardized myocardial segmentation and nomenclature for tomographic imaging of the heart. A statement for healthcare professionals from the Cardiac Imaging Committee of the Council on Clinical Cardiology of the American Heart Association. *Circulation* 105(4):539–542
40. Meloni A, Righi R, Missere M, Renne S, Schicchi N, Gamberini MR, Cuccia L, Lisi R, Spasiano A, Roberti MG, Zuccarelli A, Ait-Ali L, Festa P, Aquaro GD, Mangione M, Barra V, Positano V, Pepe A (2021) Biventricular reference values by body surface area, age, and gender in a large cohort of well-treated thalassemia major patients without heart damage using a multiparametric CMR approach. *J Magn Reson Imaging* 53(1):61–70
41. Pepe A, Positano V, Capra M, Maggio A, Lo Pinto C, Spasiano A, Forni G, Derchi G, Favilli B, Rossi G, Cracolici E, Midiri M, Lombardi M (2009) Myocardial scarring by delayed enhancement cardiovascular magnetic resonance in thalassaemia major. *Heart* 95:1688–1693
42. Meloni A, Favilli B, Positano V, Cianciulli P, Filosa A, Quarta A, D’Ascola D, Restaino G, Lombardi M, Pepe A (2009) Safety of cardiovascular magnetic resonance gadolinium chelates contrast agents in patients with hemoglobinopathies. *Haematologica* 94(11):1625–1627
43. Angelucci E, Brittenham GM, McLaren CE, Ripalti M, Baronciani D, Giardini C, Galimberti M, Polchi P, Lucarelli G (2000) Hepatic iron concentration and total body iron stores in thalassemia major. *N Engl J Med* 343(5):327–331
44. McDonagh TA, Metra M, Adamo M, Gardner RS, Baumbach A, Bohm M, Burri H, Butler J, Celutkiene J, Chioncel O, Cleland JGF, Coats AJS, Crespo-Leiro MG, Farmakis D, Gilard M, Heymans S, Hoes AW, Jaarsma T, Jankowska EA, Lainscak M, Lam CSP, Lyon AR, McMurray JJV, Mebazaa A, Mindham R, Muneretto C, Francesco Piepoli M, Price S, Rosano GMC, Ruschitzka F, Kathrine Skibelund A, Group ESCSD (2021) 2021 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure. *Eur Heart J* 42(36):3599–3726. <https://doi.org/10.1093/eurheartj/ehab368>
45. Buxton AE, Calkins H, Callans DJ, DiMarco JP, Fisher JD, Greene HL, Haines DE, Hayes DL, Heidenreich PA, Miller JM, Poppas A, Prystowsky EN, Schoenfeld MH, Zimetbaum PJ, Goff DC, Grover FL, Malenka DJ, Peterson ED, Radford MJ, Redberg RF (2006) ACC/AHA/HRS 2006 key data elements and definitions for electrophysiological studies and procedures: a report of the American College of Cardiology/American Heart Association Task Force on Clinical Data Standards (ACC/AHA/HRS Writing Committee to Develop Data Standards on Electrophysiology). *Circulation* 114(23):2534–2570
46. Cogliandro T, Derchi G, Mancuso L, Mayer MC, Pannone B, Pepe A, Pili M, Bina P, Cianciulli P, De Sanctis V, Maggio A (2008) Guideline recommendations for heart complications in thalassemia major. *J Cardiovasc Med (Hagerstown)* 9(5):515–525
47. Mos IC, Klok FA, Kroft LJ, de Roos A, Huisman MV (2012) Imaging tests in the diagnosis of pulmonary embolism. *Semin Respir Crit Care Med* 33(2):138–143. <https://doi.org/10.1055/s-0032-1311792>

48. Coutts SB (2017) Diagnosis and management of transient ischemic attack. *Continuum (Minneapolis)* 23 (1, Cerebrovascular Disease):82–92. <https://doi.org/10.1212/CON.0000000000000424>
49. Perseu L, Ristaldi MS, Dibenedetto SP, Testa R, Schiliro G, Pirastu M, Cao A (1989) The effect of the beta thalassemia mutation on the clinical severity of the sickle beta thalassemia syndrome. *Haematologica* 74(4):341–345
50. Serjeant GR, Serjeant BE, Fraser RA, Hambleton IR, Higgs DR, Kulozik AE, Donaldson A (2011) Hb S-beta-thalassemia: molecular, hematological and clinical comparisons. *Hemoglobin* 35(1):1–12. <https://doi.org/10.3109/03630269.2010.546306>
51. Benites BD, Cisneiros IS, Bastos SO, Lino A, Costa FF, Gilli SCO, Saad STO (2019) Echocardiographic abnormalities in patients with sickle cell/beta-thalassemia do not depend on the beta-thalassemia phenotype. *Hematol Transfus Cell Ther* 41(2):158–163. <https://doi.org/10.1016/j.htct.2018.09.003>
52. Poludasu S, Ramkissoon K, Salciccioli L, Kamran H, Lazar JM (2013) Left ventricular systolic function in sickle cell anemia: a meta-analysis. *J Card Fail* 19(5):333–341. <https://doi.org/10.1016/j.cardfail.2013.03.009>
53. Gladwin MT (2016) Cardiovascular complications and risk of death in sickle-cell disease. *Lancet* 387(10037):2565–2574. [https://doi.org/10.1016/S0140-6736\(16\)00647-4](https://doi.org/10.1016/S0140-6736(16)00647-4)
54. Almeida AG, Araujo F, Rego F, David C, Lopes MG, Duclasoares J (2008) Abnormal myocardial flow reserve in sickle cell disease: a myocardial contrast echocardiography study. *Echocardiography* 25(6):591–599. <https://doi.org/10.1111/j.1540-8175.2008.00666.x>
55. Acar P, Maunoury C, de Montalembert M, Dulac Y (2003) Abnormalities of myocardial perfusion in sickle cell disease in childhood: a study of myocardial scintigraphy. *Arch Mal Coeur Vaiss* 96(5):507–510
56. de Montalembert M, Maunoury C, Acar P, Brousse V, Sidi D, Lenoir G (2004) Myocardial ischaemia in children with sickle cell disease. *Arch Dis Child* 89(4):359–362. <https://doi.org/10.1136/adc.2003.027326>
57. Apostolakis S, Konstantinides S (2012) The right ventricle in health and disease: insights into physiology, pathophysiology and diagnostic management. *Cardiology* 121(4):263–273. <https://doi.org/10.1159/000338705>
58. Junqueira FP, Fernandes JL, Cunha GM, Kubo TTA, Lima CMAO, Lima DBP, Uellendhal M, Sales SR, Cunha CAS, de Pessoa VLR, Lobo CLC, Marchiori E (2013) Right and left ventricular function and myocardial scarring in adult patients with sickle cell disease: a comprehensive magnetic resonance assessment of hepatic and myocardial iron overload. *J Cardiovasc Magn Reson* 15:83. <https://doi.org/10.1186/1532-429X-15-83>
59. Mathew R, Huang J, Wu JM, Fallon JT, Gewitz MH (2016) Hematological disorders and pulmonary hypertension. *World J Cardiol* 8(12):703–718. <https://doi.org/10.4330/wjc.v8.i12.703>
60. Mushemi-Blake S, Melikyan N, Drasar E, Bhan A, Lunt A, Desai SR, Greenough A, Monaghan MJ, Thein SL, Shah AM (2015) Pulmonary haemodynamics in sickle cell disease are driven predominantly by a high-output state rather than elevated pulmonary vascular resistance. A prospective 3-dimensional echocardiography/Doppler study 10 (8). <https://doi.org/10.1371/journal.pone.0135472>
61. Ataga KI (2014) Klings ES (2014) Pulmonary hypertension in sickle cell disease: diagnosis and management. *Hematology Am Soc Hematol Educ Program* 1:425–431. <https://doi.org/10.1182/asheducation-2014.1.425>
62. Parent F, Bachir D, Inamo J, Lionnet F, Driss F, Loko G, Habibi A, Bennani S, Savale L, Adnot S, Maitre B, Yaici A, Hajji L, O'Callaghan DS, Clerson P, Girot R, Galacteros F, Simonneau G (2011) A hemodynamic study of pulmonary hypertension in sickle cell disease. *N Engl J Med* 365(1):44–53. <https://doi.org/10.1056/NEJMoa1005566>
63. Kirk P, Roughton M, Porter JB, Walker JM, Tanner MA, Patel J, Wu D, Taylor J, Westwood MA, Anderson LJ, Pennell DJ (2009) Cardiac T2* magnetic resonance for prediction of cardiac complications in thalassemia major. *Circulation* 120(20):1961–1968
64. Pepe A, Meloni A, Rossi G, Midiri M, Missere M, Valeri G, Sorrentino F, D'Ascola DG, Spasiano A, Filosa A, Cuccia L, Dello Iacono N, Forni G, Caruso V, Maggio A, Pitrolo L, Peluso A, De Marchi D, Positano V, Wood JC (2018) Prediction of cardiac complications for thalassemia major in the widespread cardiac magnetic resonance era: a prospective multicentre study by a multi-parametric approach. *Eur Heart J Cardiovasc Imaging* 19(3):299–309
65. Meloni A, Restaino G, Borsellino Z, Caruso V, Spasiano A, Zuccarelli A, Valeri G, Toia P, Salvatori C, Positano V, Midiri M, Pepe A (2014) Different patterns of myocardial iron distribution by whole-heart T2* magnetic resonance as risk markers for heart complications in thalassemia major. *Int J Cardiol* 177(3):1012–1019
66. Meloni A, Detterich J, Pepe A, Harmatz P, Coates TD, Wood JC (2015) Pulmonary hypertension in well-transfused thalassemia major patients. *Blood Cells Mol Dis* 54(2):189–194
67. Pennell DJ, Udelson JE, Arai AE, Bozkurt B, Cohen AR, Galanello R, Hoffman TM, Kiernan MS, Lerakis S, Piga A, Porter JB, Walker JM, Wood J (2013) Cardiovascular function and treatment in beta-thalassemia major: a consensus statement from the American Heart Association. *Circulation* 128(3):281–308
68. Wood JC (2008) Cardiac iron across different transfusion-dependent diseases. *Blood Rev* 22(Suppl 2):S14–21
69. Karamitsos TD, Arvanitaki A, Karvounis H, Neubauer S, Ferreira VM (2020) Myocardial tissue characterization and fibrosis by imaging. *JACC Cardiovasc Imaging* 13(5):1221–1234. <https://doi.org/10.1016/j.jcmg.2019.06.030>
70. Baker N, James J, Roy S, Wansapura J, Shanmukhappa SK, Lorenz JN, Osinska H, Backer K, Huby AC, Shrestha A, Niss O, Fleck R, Quinn CT, Taylor MD, Purevjav E, Aronow BJ, Towbin JA, Malik P (2016) Sickle cell anemia mice develop a unique cardiomyopathy with restrictive physiology. *Proc Natl Acad Sci U S A* 113(35):E5182–5191. <https://doi.org/10.1073/pnas.1600311113>
71. Niss O, Fleck R, Makue F, Alsaied T, Desai P, Towbin JA, Malik P, Taylor MD, Quinn CT (2017) Association between diffuse myocardial fibrosis and diastolic dysfunction in sickle cell anemia. *Blood* 130(2):205–213. <https://doi.org/10.1182/blood-2017-02-767624>
72. Wu J, Xia S, Kalionis B, Wan W, Sun T (2014) The role of oxidative stress and inflammation in cardiovascular aging. *Biomed Res Int* 2014:615312. <https://doi.org/10.1155/2014/615312>
73. Galie N, Humbert M, Vachiery JL, Gibbs S, Lang I, Torbicki A, Simonneau G, Peacock A, Vonk Noordegraaf A, Beghetti M, Ghofrani A, Gomez Sanchez MA, Hansmann G, Klepetko W, Lancellotti P, Matucci M, McDonagh T, Pierard LA, Trindade PT, Zompatori M, Hoeper M (2016) 2015 ESC/ERS Guidelines for the diagnosis and treatment of pulmonary hypertension. *Rev Esp Cardiol (Engl Ed)* 69(2):177. <https://doi.org/10.1016/j.rec.2016.01.002>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.